

Many Apparent False Negatives in Detection of Mutations in Gene Associated With Autism Spectrum Disorders

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Objective

To evaluate US commercial laboratories’ *SHANK3* testing quality, estimate the real-world impact of suboptimal testing on the number of undiagnosed individuals with Phelan-McDermid syndrome (PMS), and disseminate actionable findings to support accurate diagnoses of PMS

Conclusions

- False negative genetic testing results are likely being reported for individuals with ASD
- Consequently, hundreds of cases of PMS have likely gone undiagnosed in recent years, including up to half of those tested at laboratories with poor *SHANK3* testing
- These findings may inform diagnostic workflow development, motivate remedial retesting of negative results, and empower future diagnostic decision-making

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Background

- Phelan-McDermid syndrome (PMS) is a neurodevelopmental disorder that presents heterogeneously with intellectual disability (ID), speech impairment/absence, problems with social communication, motor impairments, and features of autism spectrum disorder (ASD)¹
- PMS results from disruptions to the *SHANK3* gene on chromosome 22q13, which encodes a scaffolding protein crucial for synaptic function and neuronal development²
 - While many cases of PMS involve chromosome 22q13 deletions, *SHANK3* sequence variants also commonly lead to PMS³
 - Individuals with *SHANK3* variants may have milder phenotypes⁴
- SHANK3* variants are among the most common genetic findings in ASD, affecting ~1% of individuals⁵
- Guidelines for genetic testing for neurodevelopmental disorders vary across societies, with some not reflecting current knowledge and technology⁶
- Variations in genetic testing practices among healthcare providers and the quality of genetic testing offered by diagnostic laboratories contribute to missed or delayed diagnoses
- To understand how laboratory testing quality may impact healthcare providers’ ability to accurately diagnose neurodevelopment disorders, we evaluated *SHANK3* testing quality among US diagnostic laboratories

Methods

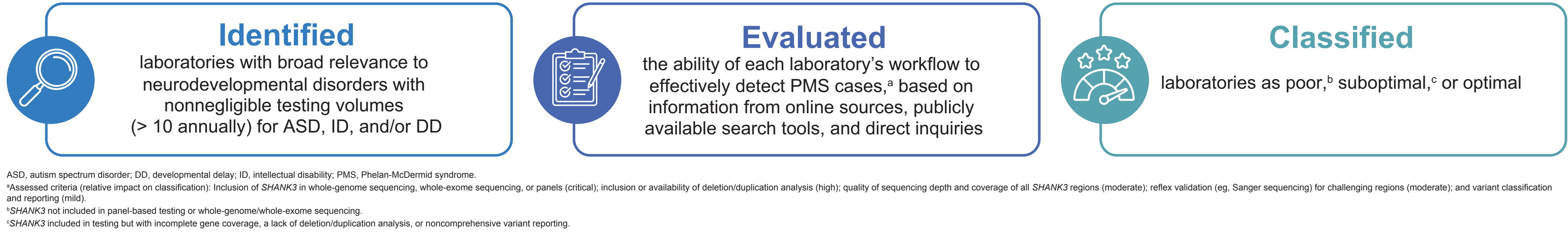
Diagnostic Laboratory Evaluation

- US diagnostic laboratories were identified, evaluated, and classified based on their next-generation sequencing (NGS) workflows (Figure 1)

Impact of Diagnostic Testing Gaps

- We used laboratory-reported testing volumes and published diagnostic yields for chromosomal microarray analysis (CMA; 7.0%) and fragile X testing (0.46%)⁷ to estimate the potential impact of the identified testing gaps at 2 laboratories on the ability to identify PMS

Figure 1. Laboratory Identification, Evaluation, and Classification



Results

Diagnostic Laboratory Evaluation

- Twelve diagnostic laboratories with relevant testing for ASD, ID, or developmental delay were identified and evaluated (Figure 2)
- NGS diagnostic methods and capabilities varied across laboratories, with significant gaps in coverage identified for several laboratories
- Identified gaps included:
 - Incomplete analysis or total omission of *SHANK3* in relevant sequencing panels
 - Lack of *SHANK3* deletion/duplication analysis
 - Reliance on exome sequencing, which may not identify large deletions
- No pattern between testing volume and quality classification was observed

Figure 2. Evaluation of Laboratories’ NGS-Based *SHANK3* Diagnostic Testing

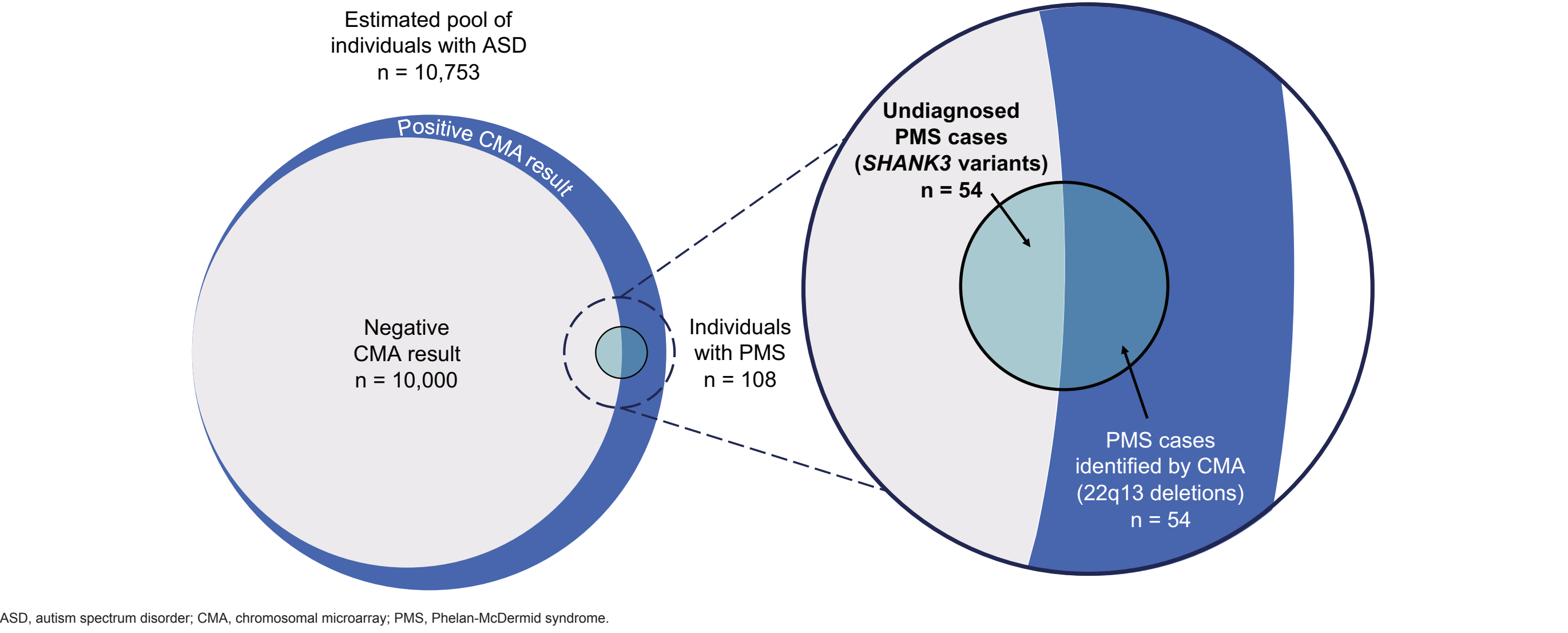
Quality Classification	Laboratory	Areas for Improvement	Annual Testing Volume
Optimal	H	Not applicable	> 10,000
	J	Not applicable	< 100
	C	Not applicable	Not available
Suboptimal	F	Relevant test does not sequence exon 11	1000–10,000
	G	Coverage of coding sequences is not guaranteed > 90%; VUS reporting is not included by default	1000–10,000
	K	Does not include deletion/duplication analysis	1000–10,000
	I	No coverage of exon 1 or portions of exon 12	100–1000
	B	Relevant panels do not include deletion/duplication analysis; exon 11 coverage is not reliable	< 100
	E	Relevant test does not include deletion/duplication analysis	Not available
Poor	D	<i>SHANK3</i> is not included in WES or relevant panels	> 10,000
	A	<i>SHANK3</i> is not included in relevant panels	< 100
	L	<i>SHANK3</i> is not included in relevant panels	Not available

NGS, next-generation sequencing; VUS, variant of uncertain significance; WES, whole-exome sequencing. Laboratories assessed as having “Optimal” quality classifications display “Not applicable” for areas of improvement.

Example Impact of Diagnostic Testing Gaps

- Laboratory D (poor testing quality) conducts > 10,000 NGS-based tests annually for individuals presenting with ASD or ID
 - Based on a CMA diagnostic yield of 7.0%,⁷ NGS tests for 10,000 individuals with ASD who had received negative CMA results would correspond to an original pool of ~10,753 individuals. Of these, 108 (1%) are likely to have PMS⁵ (Figure 3)
 - Assuming 50% of PMS cases involve large deletions (based on recent data⁸), 54 of 108 individuals with PMS would have had a 22q13 deletion identified by CMA, leaving approximately 54 individuals who would require testing with NGS methods
 - Because Laboratory D does not include *SHANK3* in NGS testing, all 54 of these individuals would remain undiagnosed

Figure 3. Potential Number of PMS Cases Missed Annually at Laboratory D



ASD, autism spectrum disorder; CMA, chromosomal microarray; PMS, Phelan-McDermid syndrome.

- NGS testing for the same 10,000 individuals with ASD at Laboratory F (suboptimal testing quality), which includes *SHANK3* but excludes exon 11 (9% of the gene’s coding sequence), would leave approximately 5 individuals undiagnosed
- Alternatively, if the NGS-based tests were ordered after fragile X testing, which has a diagnostic yield of 0.46%,⁷ the original ASD pool would comprise ~10,046 individuals, including 50 and 5 individuals with PMS undiagnosed by Laboratories D and F, respectively

Implications And Future Directions

- Based on our annual estimates of undiagnosed cases, hundreds of individuals with PMS have likely remained undiagnosed after NGS-based testing at diagnostic laboratories across the US
 - Given existing limitations to testing access, individuals receiving false negatives may not be retested
- Identifying common gaps in genetic testing can inform laboratories’ diagnostic workflow development, motivate remedial retesting of negative results, and empower decision-making by individuals, families, and healthcare providers
 - NGS-based testing for ASD, ID, and developmental delay should include *SHANK3* and copy number variant analysis to identify pathogenic or likely pathogenic variants
 - If NGS quality control metrics are low, Sanger sequencing should be performed for regions with poor coverage
 - Quality NGS-based testing can also identify PMS cases associated with deletions not detected by CMA. For example, 1 child with PMS received a negative CMA result before a whole-exome sequencing test identified an 84-kb deletion in *SHANK3* (J Holder, personal experience)
- We shared our findings with all 12 assessed laboratories
 - To date, we have received responses from 6 laboratories
 - In follow-up discussions with 3 laboratories, all expressed an interest or intent to improve; 1 laboratory classified as “poor” noted that implementing the necessary improvements could take years
- This analysis was limited to US laboratories; assessment of international diagnostic testing laboratories is needed to understand global impacts